

PCTWORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 7/01, 15/09, 15/63, 15/70, 15/74, A01N 63/00, 43/04	A1	(11) International Publication Number: WO 99/41359 (43) International Publication Date: 19 August 1999 (19.08.99)
(21) International Application Number: PCT/US99/03233 (22) International Filing Date: 16 February 1999 (16.02.99) (30) Priority Data: 60/074,844 17 February 1998 (17.02.98) US (71) Applicant: THE UAB RESEARCH FOUNDATION [US/US]; Suite 1120G/AB, 701 South 20th Street, Birmingham, AL 35294-0111 (US). (72) Inventors: CURIEL, David, T.; 824 Linwood Drive, Birm- ingham, AL 35222 (US). KRASNYKH, Victor, N.; 1832 Arboretum Circle, Birmingham, AL 35216 (US). (74) Agent: ADLER, Benjamin, A.; McGregor & Adler, LLP, 8011 Candle Lane, Houston, TX 77071 (US).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i>
(54) Title: MODIFIED ADENOVIRUS CONTAINING A FIBER REPLACEMENT PROTEIN (57) Abstract The present invention provides means to modify the tropism of recombinant adenoviral vectors using genetic methods to alter the adenoviral fiber cell-binding protein while maintaining the native trimeric protein biosynthesis profile. The present invention further provides means to specifically target particular cell types for infection with recombinant adenoviral vectors using genetic methods to alter the adenoviral fiber cell-binding protein.		

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

5

**MODIFIED ADENOVIRUS CONTAINING A FIBER
REPLACEMENT PROTEIN**

10

BACKGROUND OF THE INVENTION

Cross-Reference to Related Applications

15 This application claims benefit of U.S. provisional application Serial No. 60/074,844 filed February 17, 1998, now abandoned.

Federal Funding Legend

20 This invention was supported in part using federal funds from the National Institutes of Health. Accordingly, the Federal Government has certain rights in this invention.

Field of the Invention

25 The present invention relates generally to the fields of vector biology and gene therapy. More specifically, the present invention relates to the production of recombinant adenoviral

vectors with replacement of fibers for cell-specific targeting with concomitant elimination of endogenous tropism.

Description of the Related Art

5 Adenovirus interacts with eukaryotic cells by virtue of specific receptor recognition of domains in the knob portion of the fiber protein (21-23) which protrude from each of the twelve vertices of the icosahedral capsid. Recombinant adenovirus vectors are used in a number of gene therapy applications (21, 35,
10 38). This fact has derived principally from the high levels of gene transfer achievable with this vector approach both *in vitro* and *in vivo*.

 Recombinant adenovirus vectors are distinguished from other systems by their unique ability to accomplish *in situ*
15 gene delivery to differentiated target cells in a variety of organ contexts (5, 6, 9, 10, 12, 20, 25, 27, 29, 31). This has allowed the utilization of recombinant adenoviral vectors as an approach to treat inherited genetic diseases, such as cystic fibrosis, whereby the delivered vector may be contained within the target organ (4-
20 13). In addition, the ability of the adenoviral vector to accomplish *in situ* tumor transduction has allowed the development of a variety of anti-cancer gene therapy approaches for loco-regional disease (14-18).

 Adenoviral vectors can accomplish *in vivo* gene
25 delivery to a variety of organs after intravenous injection. In these instances, gene transfer frequencies have been sufficiently high to correct inherited metabolic abnormalities in various murine models. Thus, adenoviral vectors fulfill two requirements of an intravenously administered vector for gene therapy:

systemic stability and the ability to accomplish long-term gene expression following high efficiency transduction of muscle cells.

Adenoviruses suffer, however, from the disadvantage that the widespread distribution of the adenovirus cellular
5 receptor precludes the targeting of specific cell types. This lack of tropism of adenoviral vectors would result in a decrease in the efficiency of transduction, as the number of virus particles available for delivery to the target cells would be decreased by sequestration by nontarget cells. Furthermore, this would allow
10 ectopic expression of the delivered gene, with unknown and possibly deleterious consequences. Therefore, a means must be developed to redirect the tropism of the adenovirus vector specifically to target cells and permit gene delivery only to affected organs.

15 To this end, several groups have reported genetic modifications to the knob domain of adenovirus fiber protein and incorporation of such chimeric fibers into virion. For instance, Stevenson et al. (35) and others (24) reported successful generation of Ad5 virions containing fibers consisting of the tail
20 and shaft domains of Ad5 fiber and the knob domain of Ad3, respectively. In addition, Michael et al. (30) demonstrated the incorporation of the gastrin-releasing peptide into the carboxy terminus of recombinant Ad5 fiber. This finding was extended by Legrand et al. (30a) who achieved rescue of recombinant
25 adenovirus vectors containing such fibers. Wickham et al. (41) described the generation of recombinant virus containing fibers with carboxy-terminal polylysine sequences. These studies have established key feasibility issues with respect to this genetic approach but have also demonstrated a number of limiting factors,

including the size of the ligand, which may disrupt the correct folding of the fiber protein.

The prior art remains deficient in the lack of effective means to produce recombinant adenoviral vectors with
5 combination of novel targeting and ablation of native tropism. The present invention fulfills this longstanding need and desire in the art.

10 SUMMARY OF THE INVENTION

The present invention describes the next generation of recombinant, cell-specific adenoviral vectors. More particularly, the instant specification discloses that there are two aspects to
15 consider in the modification of adenoviral tropism: (1) ablation of endogenous tropism; and (2) introduction of novel tropism. To expand the utility of recombinant adenoviruses for gene therapy applications, methods to alter native vector tropism to achieve cell-specific transduction are necessary. To achieve such
20 targeting, one can incorporate ligands into the adenoviral fiber protein, which mediates primary binding of adenovirus to its cell surface receptor. As described herein, the present invention discloses the development of a targeted adenovirus created by replacement of the adenovirus fiber protein. The present
25 invention discloses recombinant adenovirus vectors comprising fiber replacement or substitution proteins composed of the fiber tail domain, a portion of the fibritin gene from the bacteriophage T4 and a ligand domain. The recombinant adenoviral vector may also encode a therapeutic gene i.e. the herpes simplex virus

thymidine kinase gene, in the presence of ganciclovir, the adenovirus is able to mediate the specific killing of cells which express the targeted receptor. The present invention thus represents the demonstration of the retargeting of a recombinant
5 adenoviral vector via a non-adenoviral cellular receptor.

In one embodiment of the present invention, there is provided a recombinant adenovirus vector lacking endogenous viral tropism but having novel tropism, wherein the adenovirus vector is modified to produce a replacement adenoviral fiber
10 protein so as to modify viral tropism, wherein the replaced fiber gene comprises the amino-terminal portion of the adenoviral fiber gene including the tail domain, the carboxy-terminal portion of the T4 bacteriophage fibritin gene and a ligand, wherein the replaced adenovirus fiber retains its ability to trimerize and
15 retains its native biosynthesis profile, wherein the ligand is selected from the group consisting of physiological ligands, anti-receptor antibodies and cell-specific peptides, wherein the adenoviral vector further contains a therapeutic gene, wherein said therapeutic gene is the herpes simplex virus-thymidine
20 kinase gene.

In another embodiment of the present invention, there is provided a method of killing tumor cells in an individual in need of such treatment, comprising the steps of pretreating said individual with an effective amount of the recombinant adenoviral
25 vector and administering ganciclovir to said individual.

Other and further aspects, features, and advantages of the present invention will be apparent from the following description of the presently preferred embodiments of the invention given for the purpose of disclosure.

BRIEF DESCRIPTION OF THE DRAWINGS

5 So that the matter in which the above-recited features, advantages and objects of the invention, as well as others which will become clear, are attained and can be understood in detail, more particular descriptions of the invention briefly summarized above may be had by reference to certain embodiments thereof
10 which are illustrated in the appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate preferred embodiments of the invention and therefore are not to be considered limiting in their scope.

15 Figure 1 shows the schema of the substitute fiber consisting of a fiber-fibritin-six-histidine chimera.

 Figure 2 shows a SDS gel electrophoresis of the fiber-fibritin-6histidine chimeric protein expressed in *E. coli*. Proteins were separated on 4-10% SDS-PAA gels and then stained with
20 Coomassie brilliant blue. Lane 1, fiber-fibritin-6His denatured by boiling; Lane 2, fiber-fibritin-6His native trimer; Lane 3, protein molecular mass standards.

25 DETAILED DESCRIPTION OF THE INVENTION

 The present invention is directed to an adenovirus, wherein the adenovirus is modified by replacing the adenovirus fiber protein with a fiber replacement protein. In a preferred

embodiment, the fiber replacement protein comprises: a) an amino-terminal portion comprising an adenoviral fiber tail domain; b) a chimeric fiber replacement protein; and c) a carboxy-terminal portion comprising a targeting ligand.

5 The following description will allow a person having ordinary skill in this art to determine whether a putative fiber replacement protein would function as is desired in the compositions and methods of the present invention. Generally, the fiber replacement protein associates with the penton base of
10 the adenovirus. Structurally, the fiber replacement protein is preferably a rod-like, trimeric protein. It is desirable for the diameter of the rod-like, trimeric protein to be comparable to the native fiber protein of wild type adenovirus. It is important that the fiber replacement protein retain trimerism when a sequence
15 encoding a targeting ligand is incorporated into the carboxy-terminus. In a preferred aspect, a representative example of a fiber replacement protein is T4 bacteriophage fibritin protein. More generally, the fiber replacement protein can be selected from the group consisting of trimeric structural proteins, trimeric
20 viral proteins and trimeric transcription factors. Other representative examples of fiber replacement proteins include isoleucine trimerization motif and neck region peptide from human lung surfactant D. Preferably, the fiber replacement protein has a coiled coil secondary structure. The secondary
25 structure provides stability because of multiple interchain interactions.

The fiber replacement protein does not have to be a natural protein. In fact, a person having ordinary skill in this art would be able to construct an artificial protein. Preferably, such

an artificial fiber replacement protein would have a coiled coil secondary structure.

In the adenovirus of the present invention, the targeting ligand is selected from the group consisting of physiological ligands, anti-receptor antibodies, cell-specific
5 peptides and single chain antibodies. In one embodiment, the adenovirus carries in its genome a therapeutic gene. A representative example of a therapeutic gene is a herpes simplex virus thymidine kinase gene.

10 The present invention is also directed to a method of killing tumor cells in an individual in need of such treatment, comprising the steps of: pretreating said individual with an effective amount of the adenovirus of the present invention; and administering ganciclovir to said individual.

15 Previous results obtained by modifying the Ad5 fiber suggest that the trimeric structure of the native fiber protein is not stable enough to accomodate long inserts and that the molecule will not tolerate the incorporation of large ligands. The knob seems to be the major (or only) structural component of the
20 fiber maintaining the fibers trimeric structure, while the rest of the molecule "passively" follows the trimerization process initiated by the knob. In order to create a recombinant virion lacking endogenous fiber tropism and possessing a novel tropism, one could "split" the functions of the adenoviral fiber which is
25 normally performed by the knob domain between two different protein moieties which would replace the knob. This could be achieved by replacing the knob with an external trimerization motif to maintain the trimerization of the knobless fiber, and

simultaneous introduction into the fiber of a ligand capable of targeting the virion to a novel receptor.

Since the role of the fiber is presumably to place the cell-binding site away from the surface of the virion, the fiber
5 may be replaced with another rod-like trimeric protein. In addition to being trimeric, the protein should have the ability to associate with the penton base of adenovirus and be incorporated into the virion. To prevent problems of incompatibility, the amino-terminus of the chimeric protein can be incorporated into
10 the the tail domain of the adenovirus fiber. Furthermore, the putative fiber replacement protein should tolerate the incorporation of (relatively) large additional sequences corresponding to targeting ligands into its carboxy terminus.

Fibritin, of bacteriophage T4, is the protein which
15 forms the "collar" and the "whiskers" of the phage particle and meets many of the criteria discussed above. The 486 amino acid sequence of fibritin consists of an amino-terminal domain (47 residues), a large central domain and a carboxy-terminal domain (29 residues). The central domain has 12 segments of various
20 length, each having an α -helical coiled-coil structure. Extensive biophysical and sequence analysis has modeled fibritin as a parallel triple-stranded α -helical coiled-coil. Trimers of fibritin are stable at temperatures up to 65°C. Significantly, both fibritin lacking residues at the amino terminus and recombinant fibritins
25 containing amino-terminal extensions assemble correctly into trimers. The carboxy-terminal domain of fibritin is necessary for trimerization and carboxy-terminal fusions of fibritin with the Ad5 fiber knob form stable trimers. In addition, when

overexpressed in *E.coli*, recombinant fibrin and its derivatives are soluble. Another feature of fibrin that makes it an attractive candidate for fiber replacement is the fact that it is flexible. Therefore, the length requirements established by the nature of the fiber can easily be met by fibrin, similar to the flexible fibers of bovine Ad3 and CELO virus.

Definitions

In accordance with the present invention, there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Maniatis, Fritsch & Sambrook, "Molecular Cloning: A Laboratory Manual (1982); "DNA Cloning: A Practical Approach," Volumes I and II (D.N. Glover ed. 1985); "Oligonucleotide Synthesis" (M.J. Gait ed. 1984); "Nucleic Acid Hybridization" [B.D. Hames & S.J. Higgins eds. (1985)]; "Transcription and Translation" [B.D. Hames & S.J. Higgins eds. (1984)]; "Animal Cell Culture" [R.I. Freshney, ed. (1986)]; "Immobilized Cells And Enzymes" [IRL Press, (1986)]; B. Perbal, "A Practical Guide To Molecular Cloning" (1984). Therefore, if appearing herein, the following terms shall have the definitions set out below.

A "DNA molecule" refers to the polymeric form of deoxyribonucleotides (adenine, guanine, thymine, or cytosine) in its either single stranded form, or a double-stranded helix. This term refers only to the primary and secondary structure of the molecule, and does not limit it to any particular tertiary forms. Thus, this term includes double-stranded DNA found, *inter alia*, in linear DNA molecules (e.g., restriction fragments), viruses,

plasmids, and chromosomes. In discussing the structure herein according to the normal convention of giving only the sequence in the 5' to 3' direction along the nontranscribed strand of DNA (i.e., the strand having a sequence homologous to the mRNA).

5 A "vector" is a replicon, such as plasmid, phage or cosmid, to which another DNA segment may be attached so as to bring about the replication of the attached segment. A "replicon" is any genetic element (e.g., plasmid, chromosome, virus) that functions as an autonomous unit of DNA replication *in vivo*; i.e.,
10 capable of replication under its own control. An "origin of replication" refers to those DNA sequences that participate in DNA synthesis. An "expression control sequence" is a DNA sequence that controls and regulates the transcription and translation of another DNA sequence. A coding sequence is "operably linked"
15 and "under the control" of transcriptional and translational control sequences in a cell when RNA polymerase transcribes the coding sequence into mRNA, which is then translated into the protein encoded by the coding sequence.

 In general, expression vectors containing promoter
20 sequences which facilitate the efficient transcription and translation of the inserted DNA fragment are used in connection with the host. The expression vector typically contains an origin of replication, promoter(s), terminator(s), as well as specific genes which are capable of providing phenotypic selection in
25 transformed cells. The transformed hosts can be fermented and cultured according to means known in the art to achieve optimal cell growth.

 A DNA "coding sequence" is a double-stranded DNA sequence which is transcribed and translated into a polypeptide *in*

vivo when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a start codon at the 5' (amino) terminus and a translation stop codon at the 3' (carboxyl) terminus. A coding sequence can include, but is not limited to, prokaryotic sequences, cDNA from eukaryotic mRNA, genomic DNA sequences from eukaryotic (e.g., mammalian) DNA, and even synthetic DNA sequences. A polyadenylation signal and transcription termination sequence will usually be located 3' to the coding sequence. A "cDNA" is defined as copy-DNA or complementary-DNA, and is a product of a reverse transcription reaction from an mRNA transcript.

Transcriptional and translational control sequences are DNA regulatory sequences, such as promoters, enhancers, polyadenylation signals, terminators, and the like, that provide for the expression of a coding sequence in a host cell. A "cis-element" is a nucleotide sequence, also termed a "consensus sequence" or "motif", that interacts with other proteins which can upregulate or downregulate expression of a specific gene locus. A "signal sequence" can also be included with the coding sequence. This sequence encodes a signal peptide, N-terminal to the polypeptide, that communicates to the host cell and directs the polypeptide to the appropriate cellular location. Signal sequences can be found associated with a variety of proteins native to prokaryotes and eukaryotes.

A "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a downstream (3' direction) coding sequence. For purposes of defining the present invention, the promoter sequence is bounded at its 3' terminus by the transcription initiation site

and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter sequence will be found a transcription initiation site, as well as
5 protein binding domains (consensus sequences) responsible for the binding of RNA polymerase. Eukaryotic promoters often, but not always, contain "TATA" boxes and "CAT" boxes. Prokaryotic promoters contain Shine-Dalgarno sequences in addition to the -10 and -35 consensus sequences.

10 The term "oligonucleotide" is defined as a molecule comprised of two or more deoxyribonucleotides, preferably more than three. Its exact size will depend upon many factors which, in turn, depend upon the ultimate function and use of the oligonucleotide. The term "primer" as used herein refers to an
15 oligonucleotide, whether occurring naturally as in a purified restriction digest or produced synthetically, which is capable of acting as a point of initiation of synthesis when placed under conditions in which synthesis of a primer extension product, which is complementary to a nucleic acid strand, is induced, i.e., in the
20 presence of nucleotides and an inducing agent such as a DNA polymerase and at a suitable temperature and pH. The primer may be either single-stranded or double-stranded and must be sufficiently long to prime the synthesis of the desired extension product in the presence of the inducing agent. The exact length of
25 the primer will depend upon many factors, including temperature, source of primer and use for the method. For example, for diagnostic applications, depending on the complexity of the target sequence, the oligonucleotide primer typically contains 15-25 or more nucleotides, although it may contain fewer nucleotides.

The primers herein are selected to be "substantially" complementary to different strands of a particular target DNA sequence. This means that the primers must be sufficiently complementary to hybridize with their respective strands. Therefore, the primer sequence need not reflect the exact sequence of the template. For example, a non-complementary nucleotide fragment may be attached to the 5' end of the primer, with the remainder of the primer sequence being complementary to the strand. Alternatively, non-complementary bases or longer sequences can be interspersed into the primer, provided that the primer sequence has sufficient complementarity with the sequence to hybridize therewith and thereby form the template for the synthesis of the extension product.

As used herein, the terms "restriction endonucleases" and "restriction enzymes" refer to enzymes which cut double-stranded DNA at or near a specific nucleotide sequence.

"Recombinant DNA technology" refers to techniques for uniting two heterologous DNA molecules, usually as a result of *in vitro* ligation of DNAs from different organisms. Recombinant DNA molecules are commonly produced by experiments in genetic engineering. Synonymous terms include "gene splicing", "molecular cloning" and "genetic engineering". The product of these manipulations results in a "recombinant" or "recombinant molecule".

A cell has been "transformed" or "transfected" with exogenous or heterologous DNA when such DNA has been introduced inside the cell. The transforming DNA may or may not be integrated (covalently linked) into the genome of the cell. In prokaryotes, yeast, and mammalian cells for example, the

transforming DNA may be maintained on an episomal element such as a vector or plasmid. With respect to eukaryotic cells, a stably transformed cell is one in which the transforming DNA has become integrated into a chromosome so that it is inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eukaryotic cell to establish cell lines or clones comprised of a population of daughter cells containing the transforming DNA. A "clone" is a population of cells derived from a single cell or ancestor by mitosis. A "cell line" is a clone of a primary cell that is capable of stable growth *in vitro* for many generations. An organism, such as a plant or animal, that has been transformed with exogenous DNA is termed "transgenic".

As used herein, the term "host" is meant to include not only prokaryotes but also eukaryotes such as yeast, plant and animal cells. Prokaryotic hosts may include *E. coli*, *S. typhimurium*, *Serratia marcescens* and *Bacillus subtilis*. Eukaryotic hosts include yeasts such as *Pichia pastoris*, mammalian cells and insect cells and plant cells, such as *Arabidopsis thaliana* and *Tobaccum nicotiana*.

Two DNA sequences are "substantially homologous" when at least about 75% (preferably at least about 80%, and most preferably at least about 90% or 95%) of the nucleotides match over the defined length of the DNA sequences. Sequences that are substantially homologous can be identified by comparing the sequences using standard software available in sequence data banks, or in a Southern hybridization experiment under, for example, stringent conditions as defined for that particular system. Defining appropriate hybridization conditions is within

the skill of the art. See, e.g., Maniatis et al., *supra*; DNA Cloning, Vols. I & II, *supra*; Nucleic Acid Hybridization, *supra*.

5 A "heterologous" region of the DNA construct is an identifiable segment of DNA within a larger DNA molecule that is not found in association with the larger molecule in nature. Thus, when the heterologous region encodes a mammalian gene, the gene will usually be flanked by DNA that does not flank the mammalian genomic DNA in the genome of the source organism. In another example, the coding sequence is a construct where the
10 coding sequence itself is not found in nature (e.g., a cDNA where the genomic coding sequence contains introns, or synthetic sequences having codons different than the native gene). Allelic variations or naturally-occurring mutational events do not give rise to a heterologous region of DNA as defined herein.

15 In addition, the invention may includes portions or fragments of the fiber or fibritin genes. As used herein, "fragment" or "portion" as applied to a gene or a polypeptide, will ordinarily be at least 10 residues, more typically at least 20 residues, and preferably at least 30 (e.g., 50) residues in length,
20 but less than the entire, intact sequence. Fragments of these genes can be generated by methods known to those skilled in the art, e.g., by restriction digestion of naturally occurring or recombinant fiber or fibritin genes, by recombinant DNA techniques using a vector that encodes a defined fragment of the
25 fiber or fibritin gene, or by chemical synthesis.

As used herein, "chimera" or "chimeric" refers to a single transcription unit possessing multiple components, often but not necessarily from different organisms. As used herein, "chimeric" is used to refer to tandemly arranged coding sequence

(in this case, that which usually codes for the adenovirus fiber gene) that have been genetically engineered to result in a protein possessing region corresponding to the functions or activities of the individual coding sequences.

5 The "native biosynthesis profile" of the chimeric fiber protein as used herein is defined as exhibiting correct trimerization, proper association with the adenovirus capsid, ability of the ligand to bind its target, etc. The ability of a candidate chimeric fiber-fibritin-ligand protein fragment to
10 exhibit the "native biosynthesis profile" can be assessed by methods described herein.

 A standard Northern blot assay can be used to ascertain the relative amounts of mRNA in a cell or tissue in accordance with conventional Northern hybridization techniques
15 known to those persons of ordinary skill in the art. Alternatively, a standard Southern blot assay may be used to confirm the presence and the copy number of the gene of interest in accordance with conventional Southern hybridization techniques known to those of ordinary skill in the art. Both the Northern
20 blot and Southern blot use a hybridization probe, e.g. radiolabelled cDNA or oligonucleotide of at least 20 (preferably at least 30, more preferably at least 50, and most preferably at least 100 consecutive nucleotides in length). The DNA hybridization probe can be labelled by any of the many different methods
25 known to those skilled in this art.

 The labels most commonly employed for these studies are radioactive elements, enzymes, chemicals which fluoresce when exposed to ultraviolet light, and others. A number of fluorescent materials are known and can be utilized as labels.

These include, for example, fluorescein, rhodamine, auramine, Texas Red, AMCA blue and Lucifer Yellow. A particular detecting material is anti-rabbit antibody prepared in goats and conjugated with fluorescein through an isothiocyanate. Proteins can also be
5 labeled with a radioactive element or with an enzyme. The radioactive label can be detected by any of the currently available counting procedures. The preferred isotope may be selected from ^3H , ^{14}C , ^{32}P , ^{35}S , ^{36}Cl , ^{51}Cr , ^{57}Co , ^{58}Co , ^{59}Fe , ^{90}Y , ^{125}I , ^{131}I , and ^{186}Re .

Enzyme labels are likewise useful, and can be
10 detected by any of the presently utilized colorimetric, spectrophotometric, fluorospectrophotometric, amperometric or gasometric techniques. The enzyme is conjugated to the selected particle by reaction with bridging molecules such as carbodiimides, diisocyanates, glutaraldehyde and the like. Many
15 enzymes which can be used in these procedures are known and can be utilized. The preferred are peroxidase, β -glucuronidase, β -D-glucosidase, β -D-galactosidase, urease, glucose oxidase plus peroxidase and alkaline phosphatase. U.S. Patent Nos. 3,654,090, 3,850,752, and 4,016,043 are referred to by way of example for
20 their disclosure of alternate labeling material and methods.

As used herein, the terms "fiber gene" and "fiber" refer to the gene encoding the adenovirus fiber protein. As used herein, "chimeric fiber protein" refers to a modified fiber gene as defined above.

25 As used herein the term "physiologic ligand" refers to a ligand for a cell surface receptor.

The present invention is directed to a vector system that provides both a highly efficient and specific targeting of adenovirus vector for the purpose of *in vivo* gene delivery to

predefined cell types after administration. In the recombinant adenoviral vector of the present invention, a fiber replacement protein comprising a fiber-fibritin-ligand is employed to target adenoviral vector to a specific cell for gene therapy. This is accomplished by the construction of adenoviral vectors which contain fiber-fibritin-ligand chimeras. These adenoviral vectors are capable of delivering gene products with high efficiency and specificity to cells expressing receptors which recognize the ligand component of the fiber-fibritin-ligand chimera. A person having ordinary skill in this art would recognize that one may exploit a wide variety of genes encoding e.g. receptor ligands or antibody fragments which specifically recognize cell surface proteins unique to a particular cell type to be targeted.

A "fiber replacement protein" is a protein that substitutes for fiber and provide 3 essential features: trimerizes like fiber, lacks adenoviral tropism and has novel tropism.

The following examples are given for the purpose of illustrating various embodiments of the invention and are not meant to limit the present invention in any fashion.

EXAMPLE 1

Experimental Design

To demonstrate that fibritin can be used to replace the fiber in recombinant Ad5 virions, the following experimental strategy was employed. First, a chimeric fiber-fibritin gene encoding fiber tail, the first and the second motifs of the fiber shaft domain, as well as the fiber knob domain fused with a segment of fibritin protein, was designed. In this construct, the

fibrin coding sequence replaced most of the fiber shaft coding sequence. The chimeric protein encoded by this recombinant gene will therefore resemble the Ad5 fiber protein in which almost all of the shaft domain has been replaced with the fibrin.

5

EXAMPLE 2

Construction of a recombinant adenovirus vector containing a 10 fiber-fibrin protein

In order to facilitate the subsequent transfer of the newly designed fiber gene into the Ad5 genome, the gene has been assembled in a previously made fiber shuttle vector pNEB.PK.RGD-4C. For this, pNEB.PK.RGD-4C was digested with *NaeI*
15 and *NcoI* to delete most of the fiber shaft coding sequence starting from Gly-76 through His-363. This vector was then used to clone a segment of the T4 fibrin gene encoding amino acids Ser-229 through the carboxy terminal Ala-487. The segment of the fibrin gene was PCR amplified using primers F1 (5' GGG AAC TTG
20 ACC TCA CAG AAC GTT TAT AGT CGT TTA AAT G 3') (SEQ ID No. 1) and R1 (5' AGG CCA TGG CCA ATT TTT GCC GGC GAT AAA AAG GTA G 3') (SEQ ID No. 2). In addition to the fibrin sequence, the PCR product also contained four codons at the 5' end corresponding to the fiber Gly-79 through Thr-82, whereas at the 3' end, three
25 codons corresponding to Lys-360 through Gly-362 of the fiber were added. Thus, the PCR-generated DNA fragment was digested with *NcoI* and ligated with the *NaeI-NcoI*-digested pNEB.PK.RGD-4C, resulting in pNEB.PK.FF_{BB}.

The gene assembled in pNEB.PK.FF_{BB} encodes the amino-terminal portion of the fiber protein including the complete tail domain, the first and second (and a small portion of the third) repeat of the shaft, as well as the last 222 carboxy-terminal amino acids of the fiber. The carboxy-terminal amino acids includes the entire knob, most of repeat 20 and repeats 22 and 21. The amino-terminal and carboxy-terminal portions of the fiber protein are connected by the carboxy-terminal segment of fibrin, starting from Ser-229 of the fibrin open reading frame.

In order to generate a recombinant adenovirus genome containing the fiber-fibrin gene, the plasmid pNEB.PK.FF_{BB} was used for homologous recombination with pVK50, producing pVK510. To recover the virus of interest, pVK510 was digested with *PacI* and used to transfect 293 cells. The newly generated virus, Ad5FF_{BB}, contains the fiber-fibrin protein. The generation of this virus has demonstrated that the amino-terminal portion of the fiber protein genetically fused with the fibrin sequence can efficiently associate with the Ad5 penton base, thereby allowing formation of mature virions.

EXAMPLE 3

Construction of a recombinant adenovirus vector containing a knobless fiber-fibrin protein

In order to prove the ability of the fibrin molecule to maintain the trimeric structure of the entire fiber-fibrin chimera

10

15

Efficacy of the recombinant adenovirus containing the knobless fiber-fibrin chimera

25

the chimeric protein utilizing anti-6-His tag monoclonal antibody tetra-His and anti-fiber monoclonal 4D2 which recognizes an epitope localized within the tail domain of the fiber molecule: both monoclonal antibodies identified the bands seen on the Coomassie-stained gel as fiber-fibritin-six-His chimera (data not shown).

Significantly, the fact that the fiber-fibritin chimera containing the carboxy-terminal 6-His tag efficiently bound to Ni-NTA-agarose indicates that the 6-His tag was available for binding in the context of the fiber-fibritin chimera. These results demonstrate that replacement of this tag with a targeted ligand results in an efficient interaction between a recombinant adenovirus carrying a chimeric fiber-fibritin-ligand and a ligand-specific cellular receptor.

As is well known by those having ordinary skill in this art, the fiber replacement protein will allow the incorporation of various targeting motifs e.g., targeting ligands, targeting antibodies

The following references were cited herein:

1. Jolly, D., in *Cancer Gene Therapy*, eds. Appleton & Lange, pp. 51-64, (1994).
2. Trapnell, B.C., et al., *Current Opinion in Biotechnology* 5:617-625, (1994).
3. Siegfried *Exp Clin Endocrinol* 101:7-11, (1993).
4. Bout, A., et al., *Human Gene Therapy* 5:3-10, (1994).
5. La Salle, G.L.G., et al., *Science* 259:988-990, (1993).
6. Csete, M.E., et al., *Transplantation Proceedings* 26(2):756-757, (1994).
7. Maeda, H., et al., *Gastroenterology* 106:1638-1644, (1994).
8. Jaffe, H.A., et al., *Nature Genetics* 1:372-378, (1992).

9. DeMatteo, R.P., et al., *Annals Of Surgery* 222(3):229-242, (1995).
10. Mastrangeli, A., et al., *Am J Physiol* 266:G1146-G1155, (1994).
- 5 11. Moullier, P., et al., *Kidney International* 45:1220-1225, (1994).
12. Mitani, K., et al., *Human Gene Therapy* 5:941-948, (1994).
13. Crystal, R.G., et al., *Nature Genetics* 8:42-51, (1994).
14. Clayman, G.L., et al., *Cancer Gene Therapy* 2(2):105-11, (1995).
- 10 15. Liu, T.-J., et al., *Cancer Research* 54:3662-3667, (1994).
16. Smythe, W.R., et al., *Ann Thorac Surg* 57:1395-1401, (1994).
17. Fujiwara, T., et al., *Cancer Research* 54:2287-2291, (1994).
18. Addison, C.L., et al., *Proc Natl Acad Sci* 92:8522-8526, (1995).
- 15 19. Stratford-Perricaudet, et al., *J Clin Invest* 90:626-630, (1992).
20. Huard, J., et al., *Gene Therapy* 2:107-115, (1995).
21. Henry, L.J., et al., *Journal of Virology* 68:5239-5246, (1994).
22. Stevenson, S.C., et al., *J. of Virology* 69:2850-2857, (1995).
- 20 23. Louis, N., et al., *Journal of Virology* 68:4104-4106, (1994).
24. Michael, S.I., et al., *Gene Therapy* (1995).

Any patents or publications mentioned in this specification are indicative of the levels of those skilled in the art to which the invention pertains. These patents and publications are
25 herein incorporated by reference to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and

obtain the ends and advantages mentioned, as well as those inherent therein. The present examples along with the methods, procedures, treatments, molecules, an specific compounds described herein are presently representative of preferred
5 embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention as defined by the scope of the claims.

WHAT IS CLAIMED:

1. An adenovirus, wherein said adenovirus is
5 modified by replacing the adenovirus fiber protein with a fiber replacement protein.
2. The adenovirus of claim 1, wherein said fiber
10 replacement protein comprises:
 - a) an amino-terminal portion comprising an adenoviral fiber tail domain;
 - b) a chimeric fiber replacement protein; and
 - c) a carboxy-terminal portion comprising a
15 targeting ligand.
3. The adenovirus of claim 2, wherein the fiber replacement protein associates with the penton base of the
20 adenovirus.
4. The adenovirus of claim 2, wherein said fiber replacement protein is a rod-like, trimeric protein.
- 25 5. The adenovirus of claim 4, wherein said rod-like, trimeric protein has a diameter comparable to the native fiber protein of wild type adenovirus.

6. The adenovirus of claim 2, wherein the fiber replacement protein retain trimerism when a sequence encoding a targeting ligand is incorporated into the carboxy-terminus.

5

7. The adenovirus of claim 2, wherein said fiber replacement protein is soluble.

10

8. The adenovirus of claim 2, wherein said fiber replacement protein is T4 bacteriophage fibritin protein

15

9. The adenovirus of claim 2, wherein said fiber replacement protein is selected from the group consisting of trimeric structural proteins, trimeric viral proteins and trimeric transcription factors.

20

10. The adenovirus of claim 2, wherein said fiber replacement protein is isoleucine trimerization motif.

25

11. The adenovirus of claim 2, wherein said fiber replacement protein is neck region peptide from human lung surfactant D.

12. The adenovirus of claim 2, wherein said fiber replacement protein is an artificial protein having a coiled coil secondary structure, wherein said secondary structure is stable because of multiple interchain interactions.

5

13. The adenovirus of claim 2, wherein said targeting ligand is selected from the group consisting of physiological ligands, anti-receptor antibodies, cell-specific peptides and single chain antibodies.

10

14. The adenovirus of claim 1, wherein said adenovirus carries in its genome a therapeutic gene.

15

15. The adenovirus of claim 14, wherein said therapeutic gene is a herpes simplex virus thymidine kinase gene.

20

16. A method of killing tumor cells in an individual in need of such treatment, comprising the steps of:

pretreating said individual with an effective amount of the adenovirus of claim 15; and

25

administering ganciclovir to said individual.

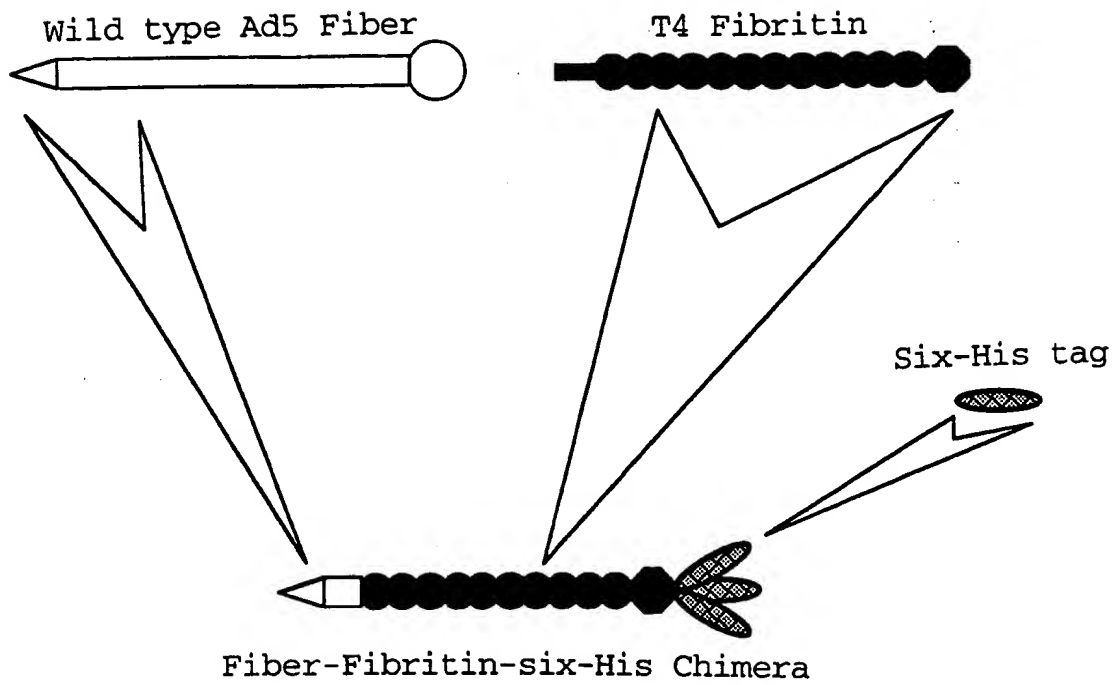


FIGURE 1

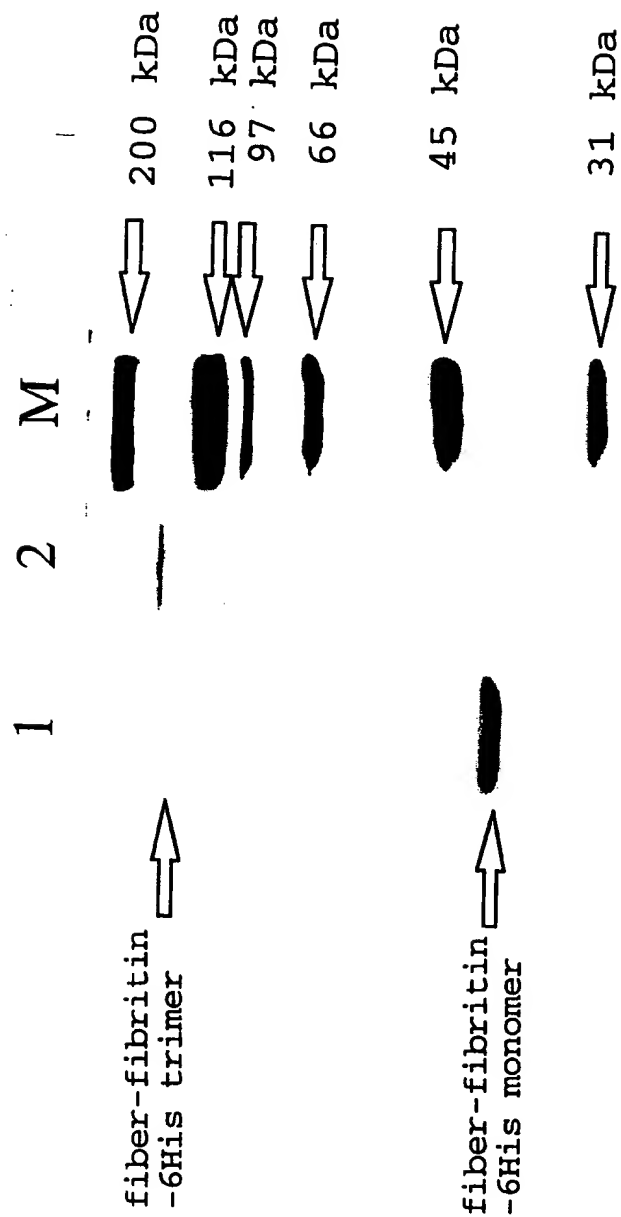


FIGURE 2

SEQUENCE LISTING

<110> Curiel, David T.
 Krasnykh, Victor N.
 <120> Modified Adenovirus Vector Containing A Fiber
 Replacement Protein
 <130> D6070PCT
 <141> 1999-02-16
 <150> US 60/074,844
 <151> 1998-02-17
 <160> 4

 <210> 1
 <211> 40
 <212> DNA
 <213> artificial sequence
 <220>
 <221> primer_bind
 <223> Forward primer F1 used to amplify segment of the T4
 fibritin gene encoding amino acids Ser-229 through
 the carboxy terminal Ala-487.
 <400> 1
 gggaacttga cctcacagaa cgtttatagt cgtttaaagt 40

 <210> 2
 <211> 37
 <212> DNA
 <213> artificial sequence
 <220>
 <221> primer_bind
 <223> Reverse primer R1 used to amplify segment of the T4
 fibritin gene encoding amino acids Ser-229 through
 the carboxy terminal Ala-487.
 <400> 2
 aggccatggc caatttttgc cggcgataaa aaggtag 37

 <210> 3
 <211> 36
 <212> DNA

<213> artificial sequence
<220>
<221> primer_bind
<223> Forward primer F2 used to amplify the 5'-terminal
segment of the fiber-fibrin gene assembled in
pNEB.PK.FF_{BB}.
<400> 3
cccctcatga agcgcgcaag accgtctgaa gatacc 36

<210> 4
<211> 42
<212> DNA
<213> artificial sequence
<220>
<221> primer_bind
<223> Reverse primer R2 used to amplify the 5'-terminal
segment of the fiber-fibrin gene assembled in
pNEB.PK.FF_{BB}.
<400> 4
ccccgatcc tgccggcgat aaaaaggtag aaagcaatac cc 42

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/03233

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C12N 7/01, 15/09, 15/63, 15/70, 15/74; A01N 63/00, 43/04

US CL : 435/235.1, 320.1; 514/44; 424/93.2, 93.6

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/235.1, 320.1; 514/44; 424/93.2, 93.6

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X — Y	US 5,543,328 A (MCCLELLAND et al) 06 August 1996, abstract and cols. 2-4.	1-3 — 4-11
X — Y	US 5,559,099 A (WICKHAM et al) 24 September 1996, abstract and cols. 2-8.	1-3 — 4-11
X — Y	US 5,712,136 A (WICKHAM et al) 27 January 1998, abstract and cols. 19-24.	1-3 — 4-11
X,P — Y,P	US 5,770,442 A (WICKHAM et al) 23 June 1998, abstract and cols. 5-10.	1-7, 9 — 8, 10, 11

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search 08 APRIL 1999	Date of mailing of the international search report 06 MAY 1999
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer BRETT L NELSON <i>JAB for</i> JOYCE BRIDGERS PARALEGAL SPECIALIST CHEMICAL MATRIX Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/03233

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P ----- Y,P	US 5,846,782 A (WICKHAM et al) 08 December 1998, abstract and cols. 13-20.	1-7, 9 ----- 9, 10, 11
X,E ----- Y,E	US 5,885,808 A (SPOONER et al) 23 March 1999, abstract and cols. 6-12.	1-7, 9 ----- 8, 10, 11
X,E ----- Y,E	US 5,871,727 A (CURIEL) 16 February 1999, abstract and cols. 8-12.	1-4 ----- 5-11

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/03233

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, DIALOG, MEDLINE, BIOSIS, EMBASE, SCISEARCH

search terms: adenovirus, recombinant, vector, fiber protein, chimera, penton, T4 bacteriophage, isoleucine trimerization, human lung surfactant